FRUCTOSAMINE AND DIABETES

H S Lim

One of the more controversial issues in the field of diabetology is whether near-normal blood glucose protects against long-term vascular complications. This is testified by the need for the on-going Diabetes Control and Complications Trial (DCCT) (1) designed to restudy the subject in IDDM.

In the meantime watching blood glucose levels remains a major preoccupation of those looking after diabetics and patients themselves. The use of glycated haemoglobin as an index of integrated glycaemic control over a period of 2-3 months has to some extent diminished the emphasis placed on isolated blood glucose readings, especially in the stable NIDDM s Its advent in the 1970s led, not surprisingly, to the discovery that many other proteins are similarly glycated by a non-enzymic posttranslational reaction to form an intermediate Schiff base which is then transformed by Amadori rearrangement into a stable ketoamine. This ketoamine end-product of protein glycation is fructosamine, the trivial name for 1-amino-1deoxyfructose. Like alvcated haemoglobin the amount of fructosamine is increased in a setting of persistent hyperglycaemia. The difference is that, because of the more rapid turn-over of circulating proteins, fructosamine reflects mean blood glucose concentration over a shorter period of 2-3 weeks.

There are at least five different ways of measuring fructosamine: (i) Phenylhydrazine Procedure (2), (ii) Furosine Procedure (3), (iii) Affinity Chromatography (4), (iv) 2-Thiobarbituric Acid (TBA) Colorimetric Procedure (5), and (v) Nitroblue Tetrazolium (NBT) Procedure (6). This last method is the subject of a paper in this issue of the SMJ.

Fructosamines are reductants under alkaline conditions. This property forms the basis for the NBT procedure in which the dye NBT is reduced to formazane which is then measured spectrophotometrically. The rate of formation of formazane is directly proportional to the fructosamine concentration. The results are expressed as mmol/L of desoxymorpholino-fructose (DMF) which is the synthetic ketoamine used as primary standard.

The NBT method is fast, simple, cheap in terms of reagents, reproducible and can be easily automated (7). A major drawback is that the test does not specifically measure glycated proteins (8). Between 25 and 60% of

Dept of Medicine IV, Tan Tock Seng Hospital. Moulmein Road Singapore 1130

H S Lim, MBBS, M Med (Int Med), Physician

Correspondence to: Dr Lim

Dept of Medicine I Singapore General Hospital Outram Road Singapore 0316

SING MED J. 1988; 29: 542 - 543

the reducing activity of fructosamine was found to be not due to non-enzymic glycation of proteins. Another problem relates to the effect of serum albumin on fructosamine values. Approximately 55% of measured fructosamine involves albumin (9). Its value is reported to be unaffected by albumin concentration above 3.0 G/L (9). Others (10) reported an effect on fructosamine over a wider range of serum albumin. Staley (11) pointed out that in the glycation of proteins, the molar concentration of reactive lysine groups of proteins is always in excess of that of carbonyl groups of glucose. The rate-limiting step is therefore the glucose concentration. Hence hypoalbuminaemia per se does not necessarily lower fructosamine concentration. Shortened half-life of proteins, as opposed to decreased synthesis, for obvious reasons, will affect fructosamine concentration. Nevertheless, various correcting factors for low albumin have been proposed (10, 12). The standardisation of the assay also presents some problems. This is because the standard, DMF, is prepared in a solution of albumin, the type and source of which affects its reactivity with NBT (7).

Currently, besides the NBT method, affinity chromatography using phenylboronic acid is the other method of fructosamine measurement that has potential for routine use by virtue of its simplicity and reported good precision (13).

The clinical usefulness of fructosamine stems from the observation that it can distinguish between normal and well-controlled diabetics from poorly-controlled patients whether it is measured by the NBT method (6), TBA method (5) or affinity chromatography (4). Further support is based on the reasonably good correlation between fructosamine and glycated haemoglobin (6) though there are exceptions (14). Good correlation is also seen between mean daily blood glucose and frustosamine (15). Within-run and day-to-day coefficients of variations (CVs) with the NBT method is typically in the range of 2-7% (16). It is gratifying that the paper in this issue of the SMJ reports CVs between 2.8 and 4.3%

These favourable observations however cannot by themselves justify widespread use of fructosamine as they apply also to glycated haemoglobin (17). What then are the situations where frustosamine could have a role? In haematological disorders such as haemolytic anaemia, iron-deficiency anaemia on treatment, and haemaglobinopathies, fructosamine may be the only reliable way to assess integrated glycaemic control. Because it reflects average blood glucose over a shorter period, fructosamine may be preferred to glycated haemoglobin in situations where patients require fortnightly or montly reviews as in the management of diabetic pregnancy. Stable patients on 3-6 monthly follow-up would benefit more from glycated haemoglobin measurements. Of course it can be argued that without a knowledge of fructosamine recent metabolic deterioration could be missed. However, such deterioration, if due to secondary failure to oral hypoglycaemic agents, seldom occurs precipitously to warrant routine fructosamine measurement. On the other hand, covert omission of insulin is likely to show

up in 2-3 weeks by a rise in fructosamine level. Hence while there is scope for its use, fructosamine should serve to complement rather than replace glycated haemoglobin, unless cost is the only consideration. At any rate, both these indices should not obviate the need for selfmonitoring of blood glucose in certain groups of patients.

Studies done so far have shown that fructosamine measurements cannot as yet replace the oral-GTT for the diagnosis of diabetes. Using the NBT method, Jury (18) found that the 95th percentile of the reference range coincided with the 10th percentile of values for diabetic subjects. The diagnostic sensitivity was only 72%. By other methods, glycated proteins and glycated albumin have been found insensitive for the detection of gestational diabetes (19). Such negative findings parallel those for glycated haemoglobin (20).

In conclusion, although fructosamine is a logical corollary of the glycated haemoglobin era and a useful parameter of medium-term glycaemic control it should be used selectively. Solving some of the problems mentioned would make it more attractive. In the final analysis, whether one measures fructosamine or glycated haemoglobin, the challenge for the laboratory is how to make its results available to the doctor "on the spot" as is the case with blood glucose. This would certainly make therapeutic decisions more precise, confident and meaningful, and perhaps reduce the frequency of outpatient visits.

REFERENCES

- 1. The DCCT Research Group: Diabetes Control and Complications Trial (DCCT): Results of Feasibility Study. Diabetes Care 1987; 10: 2-19.
- Acharya AS, Manning JM. Amadori rearrangement of glyceraldehyde haemoglobin Schiff base adducts, a new procedure for the determination of ketoamine adducts in proteins. J Biol Chem 1980; 255: 7218-24.
- Schleicher E, Scheller L, Wieland O H. Quantitation of lysine-bound glucose of normal and diabetic erythrocyte membrane by HPLC analysis of furosine. Biochem Biophys Res Comm 1981; 99: 1011-9.
- 4. Brownlee M, Vlassara H, Cerami A. Measurement of glycosylated amino acids and peptides from urine of diabetic patients using affinity chromatography. Diabetes 1980; 29: 1044-7.
- 5. McFarland KF, Catalano EW, Day JF, Thorpe SR, Baynes JW. Non-enzymatic glucosylation of serum proteins in diabetes mellitus. Diabetes 1979; 28: 1011-14.
- Johnson RN, Metcalf PA, Baker JR. Fructosamine: a new approach to the estimation of serum glycosylprotein. An index of diabetic control. Clin Chem Acta 1983; 127: 87-95.
- Baker JR, Metcalf PA, Johnson RN, Newman D, Rietz P. Use of protein-based standards in automated colorimetric determinations of fructosamine in serum. Clin Chem 1985; 31: 1550-54.
- Schleicher ED, Mayer R, Wagner EM, Gerbitz K-D. Is serum fructosamine assay specific for determination of glycated serum protein? Clin Chem 1988; 34: 320-23.
- 9. Lim YS, Staley MJ: Plasma fructosamine is a measure of all glycated proteins (Tech Brief): Clin Chem 1986; 32: 560.
- Van Dieijen-Visser MP, Seynaeve C, Burmbacher PJ. Influence of variations in albumin or total protein concentration on serum fructosamine (letter). Clin Chem 1986; 32: 1610.
- 1. Staley MJ. Fructosamine and protein concentrations in serum (letter). Clin Chem 1987; 33: 2326-27.
- 12. Howley JEA, Browning MCK, Fraser CG. Assay of serum fructosamine that minimises standardisation and matrix problems: use to assess components of biological variations. Clin Chem 1987; 33: 269-72.
- Lieper JM, Talwar D, Robb DA, Lunan CB, MacCuish AC. Glycosylated albumin and glycosylated proteins: rapidly changing indices of glycaemia in diabetic pregnancy. Q J Med 1985; 55: 225-31.
- 14. Mosca A, Carenini A, Zoppi E, et al. Plasma protein glycation as measured by fructosamine assay. Clin Chem 1987; 33: 1141-46.
- 15. Lapolla A, Poli T, Barison A, Fedele D. Fructosamine assay: an index of medium term metabolic control parameters in diabetic disease. Diab Res Clin Pract 1988; 4: 231-35.
- 16. Armbruster DA: Fructosamine: Structure, analysis and clinical usefulness. Clin Chem 1987; 33: 2153-63.
- 17. Svendson PA, Lauritzen T, Soegaard U, Nemp J. Glycosylated haemoglobin and steady-state mean blood glucose concentration in type I (insulin-dependent) diabetes. Diabetologia 1982; 23: 403-5.
- Jury DR, Dunn PJ. Laboratory assessment of a commercial kit for measuring fructosamine in serum. Clin Chem 1987; 33: 158-61.
- 19. Ryan EA, Stark R, Crockford PM, Suthijumroon A. Assessment of value of glycosylated albumin and protein in detection of gestational diabetes. Diab Care 1987; 10: 213-16.
- Little RR, England JD, Wiedmeyer HM, et al. Relationship of glycosylated haemoglobin to oral glucose tolerance. Implications for diabetes screening. Diabetes 1988; 37: 60-64.